# 1 Estimating near-infrared reflectance of vegetation from

## 2 hyperspectral data

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**Abstract:** Disentangling the individual contributions from vegetation and soil in measured canopy reflectance is a grand challenge to the remote sensing and ecophysiology communities. Since Solar Induced chlorophyll Fluorescence (SIF) is uniquely emitted from vegetation and barely influenced by the soil background, it can be used to evaluate how well reflectance-based vegetation indices (VIs) can separate the vegetation and soil components. Due to the residual soil background contributions, Near-infrared (NIR) reflectance of vegetation (NIRv) and Difference Vegetation index (DVI) present offsets as compared to SIF (i.e., the value of NIRv or DVI is non-zero when SIF is zero) when compared to SIF. In this study, we proposed a simple framework for estimating the true NIR reflectance of vegetation from Hyperspectral measurements (NIRvH) with minimal soil impacts. NIRvH takes advantage of the spectral shape variations in the red-edge region to minimize the soil effects. We evaluated the capability of NIRvH, NIRv and DVI in isolating the true NIR reflectance of vegetation using the data from both the model-based simulations and Hyperspectral Plant imaging spectrometer (HyPlant) measurements. Benchmarked by simultaneously measured SIF, NIRvH has the smallest offset (0~0.037), as compared to an intermediate offset of 0.047~0.062 from NIRv, and the largest offset of 0.089~0.112 from DVI. The magnitude of the offset can vary with different soil reflectance spectra across spatio-temporal scales, which may lead to bias in the downstream NIRv-based photosynthesis estimates. NIRvH and SIF measurements from the same sensor platform avoided complications due to different geometry, footprint and time of observation across sensors when studying the radiative transfer of reflected photons and SIF. In addition, NIRvH was primarily determined by canopy structure rather than chlorophyll content and soil brightness. Our work showcases that NIRvH is promising for retrieving canopy

structure parameters (e.g., leaf area index and leaf inclination angle) and estimating fluorescence yield with current and forthcoming hyperspectral satellite measurements.

## **Keywords:**

Solar-Induced chlorophyll Fluorescence (SIF), hyperspectral remote sensing, soil contamination, near-infrared reflectance of vegetation (NIRv), singular value decomposition (SVD), red edge

## 1.Introduction

Solar-induced chlorophyll Fluorescence (SIF) has been increasingly used to estimate the terrestrial gross primary productivity (GPP) (Frankenberg et al., 2011; Guanter et al., 2014; Ryu et al., 2019), because it originates from vegetation photosynthetic activity and is minimally influenced by soil background (Wang et al., 2019; Zeng et al., 2019). Despite these advantages, applications of SIF for GPP estimations are limited by a number of factors, including the relatively coarse spatial resolution, low revisiting frequency, low signal-noise ratio, and short history of measurements (since early 2000s) compared to the optical remote sensing (1972~) (Guanter et al., 2015). These limitations have been partly compensated with the exploitation of the newest satellites/sensors, e.g., the TROPOspheric Monitoring Instrument (TROPOMI) and Orbiting Carbon Observatory-2 (OCO-2), making a step change particularly in the spatial resolution. In the near future, the Fluorescence Explorer (FLEX) mission by the European Space Agency (Drusch et al., 2017) will provide SIF measurements at an unprecedented spatial resolution (300 m). Other limitations mentioned above will remain, including the short time series of SIF and high retrieval uncertainties.

Meanwhile, substantial knowledge on the relationship between SIF and traditional optical remote sensing has been accumulated over the recent years and can be beneficial to compensate

limitations inherent to SIF. Badgley et al. (2017), for example, proposed the near-infrared (NIR) reflectance of vegetation (NIRv) as the product of normalized difference vegetation index (NDVI) and NIR reflectance (NIRv=NIR·NDVI), which has been demonstrated as a prominent indicator of SIF under low stress conditions (Turner et al., 2020). This suggests that NIRv can be considered as the "potential SIF" (Zeng et al., 2019) and can be either used as a complement of SIF to separate the physiological and structural components (Wang et al., 2020), or as a proxy of SIF when SIF measurement is unavailable (Wu et al., 2020; Peng et al., 2020). Since NIRv is entirely based on optical surface reflectance which is much easier to acquire, it addresses many of the aforementioned limitations of SIF. Further, the availability of a "potential SIF" proxy could open new opportunities to derive SIF yield ( $\Phi_F$ ), which is a higher-level metric with even more direct linkage with vegetation photosynthesis. To date, NIRv has been used in GPP estimates, view-angle correction of SIF,  $\Phi_F$  and crop yield retrievals (Badgley et al., 2019; Hao et al., 2021a, 2021b; Peng et al., 2020; Wang et al., 2020).

However, a common limitation in traditional optical vegetation remote sensing is that observations are typically a mixture of solar radiation reflected by vegetation and the soil background. How to remove the soil contribution from observed remote sensing signals has been a challenge for optical remote sensing (Asner et al., 2002; Yang et al., 2019). As demonstrated by Badgley et al. (2017), NIRv can significantly reduce a major portion of the soil impact in the total NIR reflectance of the pixel. Zeng et al. (2019) explored the underlying physical mechanism of NIRv and found that NIRv was a good approximation of the NIR reflectance only contributed by the vegetation with a black soil background (NIR<sub>BS</sub>) and shared a similar canopy radiative transfer process as SIF (Zeng et al., 2019; Dechant et al., 2020). However, there remains residual impacts of changing soil background on NIRv (i.e., slightly violating the black-soil assumption of NIR<sub>BS</sub>), which will yield uncertainties that can propagate in the downstream NIRv applications. In fact, the varying offset (i.e., the value of a vegetation

index (VI) when SIF is zero) in the NIRv-SIF relationship due to different levels of soil contribution may complicate the universal application of NIRv as the "potential SIF" proxy, and thus its downstream applications such as estimating GPP or  $\Phi_F$  across spatiotemporal scales.

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The current formula of NIRv as NIR·NDVI can still be partially impacted by the soil background. In such a formula, NDVI serves as an adjusting factor of NIR. Ideally NDVI should range from zero (pure soil) to one (dense canopy), but in reality the soil NDVI is larger than zero, and NDVI for dense canopy is always smaller than one. Yang et al. (2020) proposed the Fluorescence Correction Vegetation Index (FCVI), which requires the NIR reflectance and the broadband visible reflectance covering the range of photosynthetically active radiation (PAR, 400~700 nm). The Difference Vegetation Index (DVI=NIR-Red; Richardson et al., 1977) was used as a proxy for FCVI (Joiner et al., 2020). DVI can also partially reduces the soil contribution due to the contrast spectra of soil and vegetation in the red-NIR region. However, DVI can still be affected by the soil background. Because soil reflectance generally increases from visible to NIR wavelengths (i.e., NIR reflectance > red reflectance), the soil DVI (=NIR-Red) is typically larger than zero and can increase with the slope of the soil visible-NIR spectrum. The sensitivity of DVI to the soil spectrum shape might be reduced by using the weighted DVI (WDVI), which puts different weights on the red and NIR bands in order to adapt to the regional soil spectrum shape, if the soil spectrum shape could be acquired in advance (Richardson et al., 1977). NDVI can be rewritten as DVI/(NIR+Red). Considering the stronger variation of the denominator than the numerator, darker soil background results in higher NDVI values, while brighter soil background leads to lower NDVI values (Qi et al., 1994; Huete et al., 2002). The increase of canopy reflectance with the wavelength is contributed by the increase of both the vegetation and soil reflectance contributions, and this effect is obvious at the red edge (e.g., 675~800 nm). With the support of current and forthcoming

hyperspectral missions, e.g., the TROPOMI covering 675~775 nm (Guanter et al., 2015), and the FLEX covering 500~780 nm (Drusch et al., 2016), it is expected to be promising to employ the red edge to better reduce the soil contribution than NIRv and DVI. For multi-spectral sensors, it is difficult to detect subtle shifts in the location and variation of the red edge reflectance driven by the chlorophyll content (Ollinger et al., 2011).

Implementing an approach that allows NIRv to be less dependent on changing soil background will open new possibilities to mitigate current limitations in SIF and enable new applications. In this study, we explored features in leaf and soil spectra shapes at the red edge to define a strategy for separating vegetation and soil contributions from measured canopy reflectance. We implemented this strategy as a new derivative of the NIR reflectance of vegetation applicable to hyperspectral data (NIRvH) with substantially reduced sensitivity to soil background, and thoroughly evaluated the proposed NIRvH against the existing NIRv and DVI approaches.

## 2. Theoretical foundation

The total bidirectional reflectance factor BRF<sub>T</sub> at wavelength  $\lambda$  of a vegetated surface can be described as a sum of three components (Zeng et al., 2019):

$$BRF_{T}(\lambda) = BRF_{BS}(\lambda) + BRF_{S}(\lambda) + BRF_{M}(\lambda)$$
 (1)

where BRF<sub>BS</sub> is the contribution of photons at wavelength  $\lambda$  reflected back by only the vegetation component over a black soil background (i.e., no soil reflection), BRF<sub>S</sub> is the single scattering contribution of photons at wavelength  $\lambda$  reflected back by the soil alone, and BRF<sub>M</sub> is the contribution of photons by multiple scattering between vegetation and soil. The goal of this study is thus to reduce the soil contribution from the total reflectance (BRF<sub>T</sub>). Whether the BRF<sub>M</sub> needs to be removed or not depends on the specific application case, and will be

discussed in Section 5.2. Based on the spectral invariants theory which considers the photon interactions with leaves and soil within the canopy in a successive order of scattering (Knyazikhin et al., 2013; Zeng et al., 2018), we can write BRF<sub>BS</sub> = $\Sigma\Sigma a_{i,j}$ · $\rho(\lambda)^i$ · $\tau(\lambda)^j$ , BRF<sub>S</sub>=b· $R_S(\lambda)$ , and BRF<sub>M</sub>= $\Sigma\Sigma\Sigma c_{i,j,k}$ · $\rho(\lambda)^i$ · $\tau(\lambda)^j$ · $R_S(\lambda)^k$ , where  $\rho(\lambda)$  and  $\tau(\lambda)$  are the leaf reflectance and transmittance at wavelength  $\lambda$ , respectively,  $R_S(\lambda)$  is the soil reflectance at wavelength  $\lambda$ , b is the bidirectional gap fraction, while  $a_{i,j}$  and  $c_{i,j,k}$  are the wavelength-independent coefficients that are within the range of 0~1 and are only determined by the canopy structure and the multiple scattering orders i, j and k. To achieve a smaller offset (the value of VI when SIF or GPP is zero) in the SIF-VI (or GPP-VI) relationship, we need to reduce the soil contribution (BRF<sub>S</sub> and part of BRF<sub>M</sub>) from the total reflectance (BRF<sub>T</sub>). The distinct features of the leaf and soil spectrum at the red edge are the key to reduce the soil contribution in BRF. Below we will describe the features of the leaf spectrum and soil spectrum at the red edge in Section 2.1 and 2.2, respectively. Then we will describe two approaches for better approximating the true NIR reflectance of vegetation with hyperspectral data in Section 2.3.

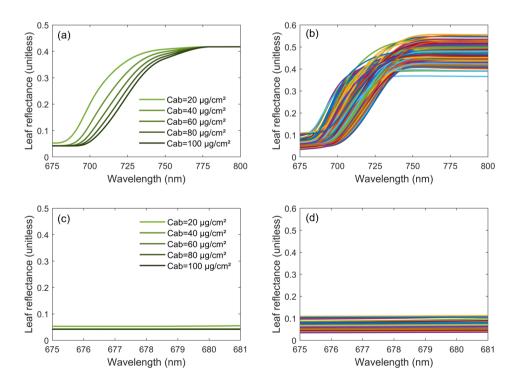
## 2.1 Leaf spectra exhibit strong absorption around 678 nm and a flat plateau in NIR

The first step to decouple the soil and vegetation contributions in total reflectance is to identify the spectral regions where the leaf spectrum is invariant while the soil spectrum is changing. There is a strong absorption feature in the leaf spectrum in the red spectral domain (675~681 nm, Fig. 1c, d), as shown in the simulations with the PROSPECT-D model and measurements in the ANGERS Leaf Optical Properties Database (Fig. 1a, b) (Feret et al., 2008; Feret et al., 2017). This feature originates from a strong absorption by chlorophyll around 675~681 nm peaking at 678 nm (Fig. S2a).

Secondly, a relatively flat plateau of the leaf spectrum within the NIR range of 778~800 nm (Figs. 1 and S1) is expected if brown pigments are not taken into consideration. The

variation of the chlorophyll content only shifts the starting and ending points of the red edge, but does not influence the reflectance in the NIR bands (Fig. 1). This is because in this wavelength region, there is little absorption of most leaf biochemical constituents, except the dry matter and brown pigments (Fig. S2). The brown pigments quantified by senescent material fraction can lead to positive slope in this wavelength region (Fig. S1b), while in practice flat plateaus are observed in the ANGERS Database (Feret et al., 2008), as shown in Fig. 1b. Overall, we find little change of the leaf spectrum in the NIR band in our modelling experiments due to the stable absorption of leaf biochemical constituents, while the brown pigments-induced uncertainties will be discussed in Section 3.1.

Due to these features, the red band (675~681 nm, Fig. 1c, d) and NIR band (778~800 nm) where the leaf spectra are relatively stable and can be used as candidate spectral regions for unmixing vegetation and soil contributions to the canopy reflectance. In practice, the selection of spectral region depends on two data features: (1) the spectral resolution, e.g., the narrow red band (675~681 nm) must have high spectral resolution (<2 nm); (2) the sensor must cover either the red region (675~681 nm) or the NIR region (778~800 nm).



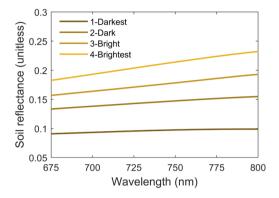
**Fig. 1** Leaf reflectance spectra simulated by the PROSPECT-D model (a, c) and measured by INRA (National Institute for Agricultural Research) from the ANGERS Leaf Optical Properties Database (Feret et al., 2008) available at <a href="http://opticleaf.ipgp.fr/index.php?page=database">http://opticleaf.ipgp.fr/index.php?page=database</a> (b, d). In the PROSPECT-D simulations, the chlorophyll content ranged from 20 μg cm<sup>-2</sup> to 100 μg cm<sup>-2</sup> at a step of 20 μg cm<sup>-2</sup>. The carotenoid content was set to 20 μg cm<sup>-2</sup>, the dry matter content was 0.012 g cm<sup>-2</sup>, the equivalent water thickness was 0.009 cm and the leaf structure parameter *N* was 1.4. The spectral range of 675~800 nm in (a) and (b) represents the red edge which we are interested in for the NIRvH estimation. The colors in (b) and (d) only represent different leaf spectra. The spectral range of 675~681 nm in (c) and (d) represents the narrow region where the leaf spectra is relatively stable in the red band.

## 2.2 Soil spectra exhibit a steady continuous change at the red edge

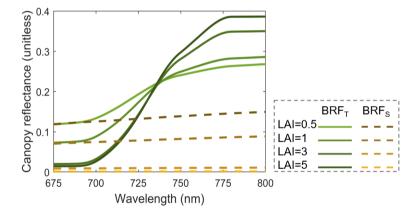
The shape of the soil reflectance spectra is typically stable with a smooth increase around the red edge from 675 nm to 800 nm (Fig. 2). Because the leaf spectrum is generally flat in red band (675~681 nm) and NIR band (778~800 nm) (Fig. 1c,d), the increase of the canopy total reflectance in the two spectral regions (Fig. 3) is primarily due to the increase of the soil reflectance. When LAI increases from 0.5 to 5 m<sup>2</sup> m<sup>-2</sup>, the contribution from soil single scattering (BRFs) to the canopy total reflectance (BRF<sub>T</sub>) may decrease, and thus the canopy spectrum becomes flatter, especially in the NIR bands. When LAI is large (e.g., 5 m<sup>2</sup> m<sup>-2</sup>), BRFs may be minimal and neglectable, and thus BRF<sub>T</sub> behaves similarly to leaf spectrum and becomes flat in the red (675~681 nm) and NIR (778~800 nm) regions. Such features offer an opportunity to extract the profile of BRFs with measurements in the two spectral regions.

In particular, we find that the red-band BRF<sub>T</sub> at the chlorophyll absorption peak (678 nm) is almost solely contributed by BRF<sub>S</sub>. For different LAIs, the vegetation contribution (BRF<sub>T</sub> - BRF<sub>S</sub>) at 678 nm does not exceed 0.02 (Fig. 3), which is even smaller than the leaf reflectance at 678 nm (generally <0.1 and mostly <0.05, Fig. 1d) considering the fractional vegetation cover, multiple scatter and strong reabsorption. Therefore, BRF<sub>T</sub> at 678 nm provides useful

information of soil reflectance. And once we know the soil spectrum profile or its slope, this information would be easy to extrapolate to other wavelengths and thereby estimate the soil contribution to BRF<sub>T</sub>.



**Fig. 2** Four soil spectrum samples with different brightness from the soil database in the SCOPE model (van der Tol et al., 2009). The spectral range of 675~800 nm in (a) and (b) represents the red edge which we are interested in for the NIRvH estimation.



**Fig. 3** Canopy-scale total reflectance (BRF<sub>T</sub>) with increasing LAI and the same soil background (dried soil, 4-Brightest in Fig. 2), simulated by the SCOPE model. The soil single scattering contribution (BRF<sub>S</sub>) is calculated by the product of the bidirectional gap fraction times the soil reflectance. The spectral range of 675~800 nm represents the red edge which we are interested in for the NIRvH estimation.

## 2.3 Two approaches to develop the hyperspectral NIRv (NIRvH)

Based on the analysis above, we developed two simple and practical approaches (i.e., NIRvH1 and NIRvH2) for the NIRvH index without using any additional canopy structure parameters. In principle, BRF<sub>T</sub> can be written in another way as the sum of vegetation and soil contributions (BRF<sub>Veg</sub> and BRF<sub>Soil</sub>, respectively):

$$BRF_{T}(\lambda) = BRF_{Veg}(\lambda) + BRF_{Soil}(\lambda)$$
 (2)

NIRvH1 is a generalized approach that uses a singular value decomposition (SVD) method to estimate BRF<sub>Soil</sub> and a logistic function to fit the BRF from vegetation (BRF<sub>Veg</sub>) at the red edge (675~800 nm). Compared to Eq. (1), BRF<sub>Soil</sub> is the sum of BRF<sub>S</sub> and a part of BRF<sub>M</sub>, while BRF<sub>Veg</sub> includes BRF<sub>BS</sub> and the remaining portion of BRF<sub>M</sub> in BRF<sub>T</sub>.

BRF<sub>Soil</sub> at the red edge can be estimated as

$$BRF_{Soil}(\lambda) = \sum \beta_i \cdot PC_i(\lambda)$$
 (3)

where  $PC_i$  are the principal components extracted from the neighbouring pure soil pixels in the same image or from the soil spectral database, and  $\beta_i$  are the fitted coefficients. The shape of soil spectrum at the red edge within 675~800 nm is typically as simple as a linear line (Fig. 2). The first one or two PCs usually can explain more than 95% of the soil spectrum variance, and the complexity of the soil spectrum shape determines the number of PCs needed for the fitting. With the SVD method, we do not need to assume the shape of the soil spectrum is linear at the red edge.

BRF<sub>Veg</sub> at the red edge can be approximated as a logistic function

BRF<sub>Veg</sub>(
$$\lambda$$
)=a/(1+b·exp(-t·( $\lambda$ - $\lambda_0$ ))) (4)

253 where a, b and t are the fitted coefficients, and  $\lambda_0$  is 675 nm where the canopy spectrum begins 254 to increase.

Combining Eqs. X and Y gives

$$BRF_{T}(\lambda) = BRF_{Veg}(\lambda) + BRF_{Soil}(\lambda) = \sum \beta_{i} PC_{i}(\lambda) + a/(1 + b \cdot exp(-t \cdot (\lambda - \lambda_{0})))$$
 (5)

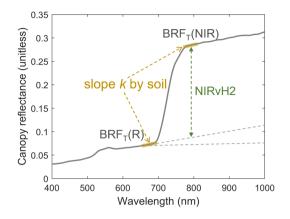
Therefore, we can use the canopy reflectance to fit Eq. 5 to determine the coefficients ( $\beta_i$ , 258 a, b, and t), and thus directly estimate NIRvH1 using Eq. 4 and the fitted a, b, t and  $\lambda$  in the NIR band (e.g., 778 nm).

Considering the delicate atmospheric correction of high spectral resolution data, using the "atmospheric windows" is a more pragmatic strategy to avoid these errors. Therefore, we used the wavelengths at the "atmospheric windows" instead of the full spectral range in our NIRvH1 approach.

NIRvH2 was developed by assuming linear soil spectra at the red edge (Wang et al., 2017; Yang et al., 2019). Although this assumption might not be universally accurate, it could be reasonable for the most common cases. With this assumption, NIRvH2 can be written as

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$$NIRvH2 = BRF_T(NIR) - BRF_T(R) - k \cdot (\lambda_{NIR} - \lambda_R)$$
 (6)

where BRF<sub>T</sub>(R) and BRF<sub>T</sub>(NIR) are the reflectance in the red and NIR bands, respectively (Fig. 4). According to the typical shapes of soil and vegetation spectrum shapes,  $\lambda_R$  is set to be 678 nm, while  $\lambda_{NIR}$  could be within 778~800 nm for the best performance. BRF<sub>T</sub>(678 nm) is contributed mostly by BRF<sub>S</sub>(678 nm) (the product of the bidirectional gap fraction 'b' and the soil reflectance  $R_S$ (678 nm)) in Eq. 1, and thus by the fitting with the baseline reflectance BRF<sub>T</sub>(678 nm) in Fig. 4, we do not need to know either 'b' or  $R_S$ (678 nm), separately. The slope parameter, k, can be estimated by linear fitting several BRF<sub>T</sub> against wavelengths in either one of the two leaf spectral-invariant regions: red band (675~681 nm) or NIR band (778~800 nm) as discussed in Section 2.1, depending on the data availability. Compared to NIRvH1, NIRvH2 is simpler and does not require the neighbouring pure soil pixels or a soil spectral database to extract the PCs. Hyperspectral reflectance is not necessary for NIRvH2 because the slope k can be derived from a few multispectral bands in the two spectral regions. However, the performance of NIRvH2 depends on linearity of the soil spectrum at the red edge, and thus has a narrower range of applications than NIRvH1.



**Fig. 4** A diagram demonstrating the calculation of the NIRvH2 index from the canopy total reflectance by the baseline reflectance BRF<sub>T</sub>(R) at 678 nm, and by the slope k (Eq. 6) from either the red region (675~681 nm) or the NIR region (778~800 nm).

## 3. Materials and methods

#### 3.1 Model-based evaluation with the SCOPE model

The Soil Canopy Observation, Photochemistry and Energy (SCOPE) model version 1.70 (van der Tol et al., 2009) was used to simulate canopy reflectance and SIF with various combinations of canopy structure, leaf property, sun-sensor geometry and soil spectrum (Table 1), in order to test the performance of NIRvH1 and NIRvH2. Four soil spectra with different brightness levels from the soil database in the SCOPE model (Fig. 2) were used in the simulations. All the other parameters were kept as the default values in SCOPE v1.70. In total, 20,736 different combinations were generated. The broadband incoming shortwave radiation (400-2500 nm) was set at 600 W m<sup>-2</sup>. The simulated reflectance covered 400~1000 nm with a sampling interval of 1 nm, and simulated SIF at the O<sub>2</sub>-A band (760 nm) was used for the analysis. NIR<sub>BS</sub> was obtained by replacing the soil with a black-body background in the model setup. We chose the central wavelengths of the Moderate Resolution Imaging

Spectroradiometer (MODIS) sensor at the red band (648 nm) and NIR band (858 nm) (Vermote et al., 1997) to calculate NIRv and DVI.

In order to evaluate the practical performance of NIRvH1 and NIRvH2, we chose to use the spectral range of 675~775 nm to be identical to the spectral coverage of existing hyperspectral satellite sensors, e.g., TROPOMI. The four soil spectra in Fig. 3 were used in the SVD method for the NIRvH1 calculation. For calculating NIRvH2, we set  $\lambda_{NIR}$ =775 nm. The slope parameter k in Eq. 6 was estimated using the simulated red band (675~681 nm) canopy reflectance to be consistent with TROPOMI.

We compared four simulated VIs (NIRvH1, NIRvH2, DVI and NIRv) to evaluate their performance in reducing the soil impacts. To highlight the conditions of sparse vegetation, we first examined the results where SIF was no more than 1 W m<sup>-2</sup> um<sup>-1</sup> sr<sup>-1</sup> and all the VIs were no more than 0.3. Both SIF and NIR<sub>BS</sub>, which had little soil influence, were used as the benchmark to evaluate the soil impacts on the four VIs. The offsets (i.e., the value of a vegetation index when SIF (or NIR<sub>BS</sub>) is zero) in the VI-SIF (or VI-NIR<sub>BS</sub>) relationship, were used to indicate the impact of soil brightness on VIs. For example, if the linear fitting line between a VI and SIF or NIR<sub>BS</sub> goes through the origin point of coordinates (0, 0), the offset is zero and suggests promising performance of the VI in reducing the soil influence. In addition, the relative offset (RO) was also calculated as the ratio between the offset and the mean of the VI. Similar analyses were also conducted with the model simulations where NIRvH1 and NIRvH2 were larger than 0.3 for the evaluation over dense canopies.

As suggested in several earlier studies (Zeng et al., 2019; Yang et al., 2020; Hao et al., 2021a), pure vegetation reflectance has a similar sun-sensory geometry effects as SIF and thus could be used to correct the anisotropy of SIF. Therefore, a VI that can well approximate pure vegetation reflectance should demonstrate good performance on correcting the anisotropy of SIF observations at different sun-sensor geometries. With the setting of varied sun-sensor

geometry in our SCOPE simulations, we evaluated the four VIs for their ability of correcting the anisotropy of SIF in two approaches. One approach was to calculate the total emitted SIF by all leaves (SIF<sub>Total</sub>) from top-of-canopy SIF (SIF<sub>TOC</sub>) with different sun-sensor geometries: SIF<sub>Total</sub> = SIF<sub>TOC</sub>/fesc, and compared it with the true value provided by model simulation. SIF<sub>Total</sub> can be estimated from SIF<sub>TOC</sub> obtained under a sun-sensor geometry by dividing SIF<sub>TOC</sub> by a photon escape ratio  $fesc = VI/(fPAR \cdot w)$ , where VI is one of the four abovementioned VIs under the same sun-sensor geometry of SIFTOC, fPAR is the fractional of absorbed photosynthetically active radiation (PAR) from SCOPE in this study, and w is the leaf single scattering albedo at 760 nm (Zeng et al., 2019; Dechant et al., 2020). The second approach was to normalize SIF<sub>TOC</sub> to the nadir viewing direction (SIF<sub>Nadir</sub>), with the corresponding VIs at the same angle of SIF<sub>TOC</sub> (by VI<sub>TOC</sub>) and nadir VIs (VI<sub>Nadir</sub>) (Zeng et al., 2019; Hao et al., 2021a, 2021b): SIF<sub>Nadir</sub>=SIF<sub>TOC</sub>\*VI<sub>Nadir</sub>/VI<sub>TOC</sub>. The calculated SIF<sub>Nadir</sub> can be compared with the true value provided by model simulations to evaluate the anisotropy of VI in comparison to SIF. The first approach may have systematic biases due to different behaviours of SIF and VI over the bare soil, because VI may not be equal to zero when SIF was zero. The second approach highly depends on the anisotropy distribution of SIF and VI, while the soil impacts on VIs could vary with the sun-sensor geometry with different fractions of sunlit/shaded soil in view.

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**Table 1**. List of variables and their ranges used in SCOPE v1.70 simulations of canopy reflectance and fluorescence (Zeng et al., 2019), resulted in 20,736 combinations. Default values were used for all the other variables.

	Variable	Values
Canopy Structure	Leaf Area Index (LAI, m <sup>2</sup> m <sup>-2</sup> )	[0.5, 1, 3, 5]
	Leaf Angle Distribution	Spherical, Erectophile, Planophile

Leaf Property	Leaf Chlorophyll Content (Cab, μg cm <sup>-2</sup> )	[40, 60, 80]
	Dry Matter Content (Cm, g cm <sup>-2</sup> )	[0, 0.025, 0.05]
	Brown Pigments (Senescent material fraction, Cs)	[0, 0.3, 0.6, 0.9]
Sun-sensor Geometry	Solar Zenith Angle	[30°, 50°]
	View Zenith Angle	[0°, 30°, 50°]
	Relative Azimuth Angle	[0°, 180°]
Soil Background Soil Spectra		Four soil spectra

### 3.2 HyPlant airborne dataset

We also used hyperspectral and SIF measurements from an airborne remote sensing experiment with the HyPlant sensor to evaluate NIRvH1 and NIRvH2. The HyPlant sensor (Specim, Finland) is an airborne imaging spectrometer for vegetation monitoring with two sensors operating in push-broom mode (Rascher et al., 2015; Siegmann et al., 2019). The fluorescence imager (FLUO) module of HyPlant acquires radiance at high spectral resolution (0.25 nm) in the spectral region of the two oxygen absorption bands (670~780 nm), enabling simultaneous acquisition of SIF and reflectance with consistent geometry. The DUAL module of HyPlant provides imageries covering the spectral range of 380~2500 nm with the spectral resolution at about 1.7 nm at the red edge. In this study, four HyPlant airborne imageries were acquired with the flight altitude of 600 m above the ground surface, including three regions of agricultural fields (Region A~C), and one region of complex land surfaces (Region D). The imageries were acquired on August 23<sup>rd</sup>, 2012 (Region A: 50°52'37.52"N/ 6°26'41.89"E, sugar beet), June 30<sup>th</sup>, 2015 (Region B: 50°52'33.42"N/ 6°26'10.83"E, potato, winter wheat and sugar beet), July 19<sup>th</sup>, 2016 (Region C: 50°52'43.95"N/ 6°26'11.79"E, sugar beet, potato

and maize), and August 23<sup>rd</sup>, 2012 (Region D: 50°54'39.10"N/ 6°26'41.89"E, forest with woody components, water body and urban areas), respectively (Fig. S3).

The improved Fraunhofer Line Discrimination (iFLD) module was adopted to retrieve SIF at the O<sub>2</sub>-A band (760 nm) from the observed radiance (Damm et al., 2011). The HyPlant imageries were atmospherically corrected using the MODTRAN radiative transfer model to obtain the relevant atmospheric transfer functions and enable the calculation of surface reflectance (Berk et al., 2005). The spatial resolution for reflectance and SIF was originally at 1 m but was aggregated to 5 m to reduce the noise and compensate the impact of a wide spatialspectral point spread function. For NIRvH1, we chose the same wavelength of TROPOMI covering 675~775m. Again, only the wavelengths at the atmospheric windows within 675~775 nm of the HyPlant FLUO surface reflectance data were used for the NIRvH1 (cf. Section 2.3 for a justification). The soil pixels for the SVD method in NIRvH1 were selected by a criterion which has a threshold value of 0.2 for NDVI at the same HyPlant imagery. For NIRvH2, we set  $\lambda_{NIR}$ =775 nm and extracted k in Eq. 6 with linear fitting of the total reflectance within the range of 675~681 nm or 778~800 nm. Because NIR<sub>BS</sub> cannot be directly acquired by the airborne data, we only used far-red SIF at 760 nm as the reference. NIRv and DVI were also included in the HyPlant analysis to compare with NIRvH1 and NIRvH2. Reflectance at 648 nm (red) and 858 nm (NIR), which were identical to the central wavelengths of MODIS, were used to calculate NIRv and DVI.

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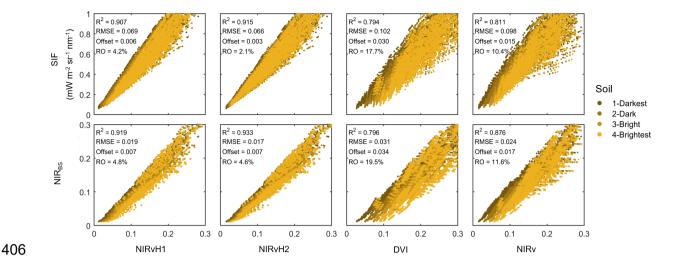
## 4. Results

#### 4.1 Evaluation by SCOPE simulations

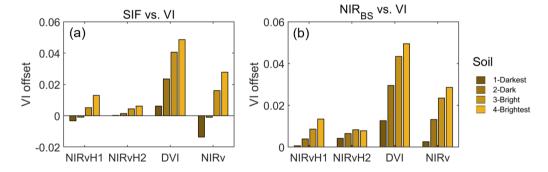
All the four VIs demonstrated strong linear relationships with SIF and NIR<sub>BS</sub>, and the coefficient of determination ( $R^2$ ) was no less than 0.79 (Fig. 5) by the SCOPE simulations at

different combinations of canopy structure, leaf property, sun-sensor geometry and soil optical properties in Table 1. NIRvH1 and NIRvH2 stood out and showed the strongest and similar correlations with SIF and NIR<sub>BS</sub>, with the highest  $R^2$  and lowest RMSE, as well as the smallest offset and RO. In contrast to the converged small offsets of NIRvH1 and NIRvH2, DVI and NIRv had various and larger offsets in the SIF-VI or NIR<sub>BS</sub>-VI relationships in response to the four soil spectra with different levels of brightness in the SCOPE simulations. ROs of DVI and NIRv were also higher (Fig. 5). In the SIF-VI relationship, ROs were 17.7% and 10.4% for DVI and NIRv, while they were 4.2% and 2.1% for NIRvH1 and NIRvH2, respectively. Similarly, in the NIR<sub>BS</sub>-VI relationship, ROs were 19.5% and 11.6% for DVI and NIRv, and were 4.8% and 4.6% for NIRvH1 and NIRvH2, respectively.

Larger offsets were found for DVI and NIRv (Figs. 5~6) with brighter soil backgrounds, which result in steeper red-edge slopes in the TOC reflectance spectra (Fig. 2). The largest offset over the brightest soil in the SIF-VI relationship for DVI and NIRv could be as large as 0.049 and 0.028, respectively, while the largest offset for NIRvH1 or NIRvH2 was generally less than 0.013 (Fig. 6a). The largest offset over the brightest soil in the NIR<sub>BS</sub>-VI relationship for NIRvH1 and NIRvH2 was 0.013 and 0.008, and was 0.029 and 0.050 for NIRv and DVI, respectively (Fig. 6b). Over dense canopies when all the four VIs were larger than 0.3, NIRvH1 and NIRvH2 still performed better than DVI and NIRv in approximating NIR<sub>BS</sub>, with a slightly higher  $R^2$  and lower RMSE when compared to NIR<sub>BS</sub> (Fig. S4).



**Fig. 5** Scatter plots between SIF at 760 nm or NIR<sub>BS</sub> and different VIs by SCOPE simulations at different combinations of canopy structure, leaf property, sun-sensor geometry and soil spectrum in Table 1. To highlight the condition of sparse vegetation, we first examined the results where SIF was no more than 1 W m<sup>-2</sup> um<sup>-1</sup> sr<sup>-1</sup> and all the four VIs were no more than 0.3. Note there were four soil spectra in the simulation, which led to four different offsets for the SIF-VI or NIR<sub>BS</sub>-VI relationships for DVI and NIRv as in Fig. 6. The relative offset (RO) was estimated by the ratio of the offset and the mean of the corresponding VI.



**Fig. 6** Offsets in the relationship between SIF at 760 nm or NIR<sub>BS</sub> with different VIs (Fig. 4) over four different soil brightness by SCOPE simulations. The simulations were conducted with different combinations of canopy structure, leaf property, sun-sensor geometry and soil reflectance spectra in Table 1.

All the four VIs demonstrated good performances in the SIF anisotropy correcting experiments with SCOPE, i.e., the  $R^2$ s between the corrected and simulated data were no lower than 0.78 (Fig. 7). NIRvH1 and NIRvH2 had higher  $R^2$ s and lower RMSEs than DVI and NIRv for both SIF<sub>Total</sub> and SIF<sub>Nadir</sub>. For sparse vegetation canopies (e.g., LAI=0.5 m<sup>2</sup> m<sup>-2</sup>), using DVI

and NIRv may result in underestimated SIF<sub>Total</sub> by over 30% and 10%, respectively (Fig. 8). This is due to the overestimations of *fesc* by DVI and NIRv, caused by the residual soil effects. SIF<sub>Nadir</sub> using DVI and NIRv were overestimated by about 20% and 10% at maximum, again due to the residual soil contributions that led to different anisotropic distributions between SIF and DVI or NIRv. The residual soil contributions to DVI or NIRv changed with the sun-sensor geometry, caused by different fractions of sunlit/shaded soil in view. In contrast, we found the relative error was less than 10% for both SIF<sub>Total</sub> and SIF<sub>Nadir</sub> over all the sun-sensor geometries if NIRvH1 or NIRvH2 were used.



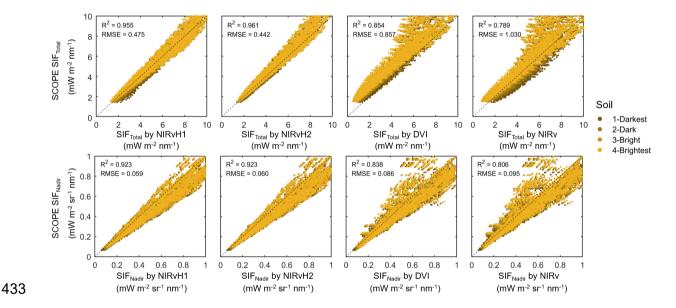
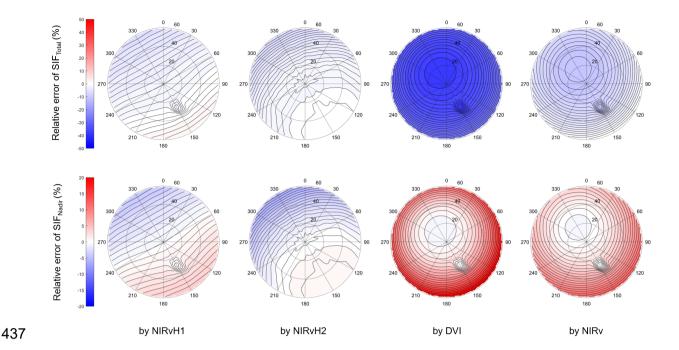


Fig. 7 Scatter plots between  $SIF_{Total}$  or  $SIF_{Nadir}$  at 760 nm normalized by different VIs and SCOPE references at different combinations of canopy structure, leaf property, sun-sensor geometry and soil spectrum in Table 1.

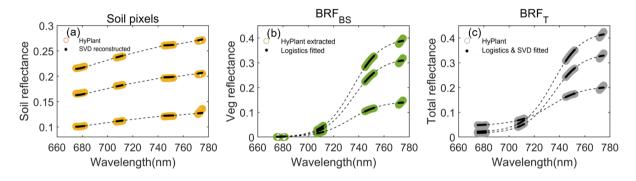


**Fig. 8** Relative error of SIF<sub>Total</sub> or SIF<sub>Nadir</sub> normalized by different VIs over sparse canopies when compared to SCOPE references at different viewing geometries, with the solar zenith angle at 30°, the leaf area index at 0.5 m<sup>2</sup> m<sup>-2</sup>, the same soil background (dried soil, 4-Brightest in Fig. 2), spherical leaf angle distribution, the chlorophyll content at 60 μg cm<sup>-2</sup>, the carotenoid content at 20 μg cm<sup>-2</sup>, the dry matter content at 0.025 g cm<sup>-2</sup>, the equivalent water thickness at 0.009 cm, the senescent material fraction at 0.3, and the leaf structure parameter N at 1.4. The contour lines divide each plot into equal intervals, and the color from blue to red represents the relative error from negative to positive.

#### 4.2 Evaluation by airborne HyPlant dataset

The performance of the NIRvH1 calculation on the spectral fitting by the SVD method and the logistic function at three spectra from the HyPlant imagery was displayed in Fig. 9. The displayed three spectra of soil or total reflectance were sampled at 20%, 50%, and 80% percentiles of the data (Fig. 9). The increase of HyPlant-measured soil reflectance was not strictly linear at the red edge (675~775 nm). The first *PC* by the SVD method explained 98% of the soil spectrum variance for the pure soil pixels at region A, and only the first *PC* was thus used in the fitting. The results show that the SVD-reconstructed soil reflectance agreed well with the HyPlant-measured soil reflectance (Fig. 9a). After the removal of the soil contribution

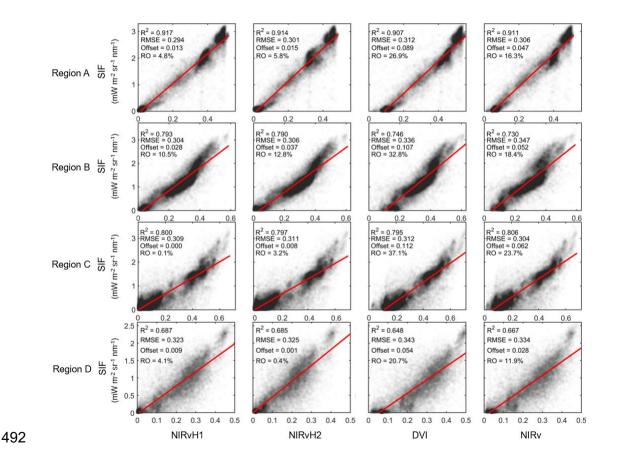
by the SVD method, the remaining HyPlant vegetation reflectance of the three pixels agreed well with the shape of the leaf spectrum, and can be well fitted by a logistic function (Fig. 9b). Overall, the HyPlant total reflectance of the three pixels with different soil brightness as shown in the red band (Fig. 9c), can be well fitted by the SVD method and the logistic function in the NIRvH1 calculation.



**Fig. 9** The performance of the fitting by the singular value decomposition (SVD) method and the logistic function in the NIRvH1 calculation at three spectra from the HyPlant fluorescence imager at region A in Fig. S3. (a) The HyPlant measured reflectance of three soil pixels representing the 20%, 50%, and 80% percentiles of the soil reflectance values, and the corresponding SVD reconstructed reflectance. (b) The extracted HyPlant vegetation reflectance (BRF<sub>BS</sub>) after the removal of the soil contribution by the SVD approach at three mixed pixels representing the 20%, 50%, and 80% percentiles of the total reflectance values, and the corresponding logistic function-fitted vegetation reflectance. (c) The HyPlant observed total reflectance (BRF<sub>T</sub>) at the three mixed pixels, and the corresponding fitted total reflectance by the SVD method and the logistic function in the NIRvH1 calculation. Note only the wavelengths in the atmospheric windows with high transmittance around 680 nm, 710 nm, 750 nm and 775 nm were used.

Scatter plots in Fig. 10 show the comparison between HyPlant-measured SIF and different Vis. In the three agricultural field regions (region A $\sim$ C), all the four VIs showed strong linear correlations with SIF ( $R^2 > 0.73$ ). Positive offsets were found for all the four VIs in the SIF-VI relationship over sparse vegetation canopies. DVI and NIRv had higher ROs at the levels of 26.9% $\sim$ 37.1% and 16.3% $\sim$ 23.7%, respectively. In contrast, ROs for NIRvH1 and NIRvH2

were only 0.1%~10.5% and 3.2%~12.8%, respectively. NIRvH1 and NIRvH2 had the smallest offsets within 0~0.028 and 0.008~0.037, respectively. NIRvH1 performed slightly better than NIRvH2 because the shape of soil reflectance spectra might be not strictly linear (Fig. 10a). NIRv had an intermediate offset within 0.047~0.062, and DVI had the largest offsets ranged between 0.089 and 0.112, two to three folds higher than NIRvH1 and NIRvH2. In the region D with more complex land surfaces, NIRvH1 and NIRvH2 kept showing better performance than DVI and NIRv, with higher  $R^2$ , lower RMSE, offset and ROs. ROs for NIRvH1 and NIRvH2 were only 4.1% and 0.4%, respectively, while for DVI and NIRv were as large as 20.7% and 11.9%, respectively. Overall, Fig. 10 suggests that the traditional DVI and NIRv are not promising at removing soil impacts with non-zero offset in their linear regression line with SIF, which may be primarily due to the increasing shape of soil reflectance at the red edge. By contrast, the soil effect was largely reduced with the proposed NIRvH1 and NIRvH2.



**Fig. 10** Scatter plots of SIF at 760 nm and different VIs for HyPlant imageries at four study regions on August 23<sup>rd</sup>, 2012, June 30<sup>th</sup>, 2015, July 19<sup>th</sup>, 2016 and August 23<sup>rd</sup>, 2012, respectively. Regions A~C were agricultural fields, while region D was over complex land surfaces composed of forest, water body and urban areas. The relative offset (RO) was estimated by the ratio of the offset to the mean value of the corresponding VI in the SIF-VI relationship.

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## 5. Discussion

## 5.1 Towards better separation of soil and vegetation contributions in canopy reflectance

## by NIRvH

To date, NIRv and DVI have been considered as effective approaches of minimizing soil impacts and widely used in measuring vegetation growing status, and this may be of particular concern when they are used to reduce structural effects inherent to SIF retrievals (Zeng et al., 2019; Yang et al., 2020; Joiner et al., 2020). Here we found that they may have varying positive offsets when compared to SIF and BRF<sub>BS</sub>, i.e., positive values at bare soil condition. This is mainly caused by the increasing soil reflectance at the red edge (Figs. 5 and 9), and thus soil still partly contributes to the difference between NIR and red reflectance, which was not fully accounted in their mathematical formulas. Furthermore, different sensor settings may introduce an additional level of complexity due to sampling red and NIR reflectance at different spectral wavelengths (e.g., MODIS at 648 nm and 858 nm, TROPOMI at 675 nm and 775 nm for red and NIR central wavelength, respectively) which are associated with different soil reflectance. Sellers et al (1992) emphasized the importance of disentangling the individual contributions from vegetation and soil in the red and NIR, and that this was particularly challenging with multispectral data especially over sparse vegetation canopies. We made full use of the shape of the soil and leaf spectrum at the red edge covered by hyperspectral data, and developed the NIRvH1 and NIRvH2. Both indices show strong linear correlations with SIF and BRF<sub>BS</sub> (Figs.

5 and 10), and can be directly used for the angular correction of directional observed SIF with good performance (Figs. 7 and 8). These two indices are promising for improving recent studies with NIRv, especially over sparse vegetation canopies at agricultural, forest and urban ecosystems (Fig. 10). While the goals of reducing the soil contribution in BRF can also be achieved with a physically-based canopy radiative transfer model, it requires retrievals of soil and vegetation structural parameters (e.g., leaf area index and leaf angle distribution) and thus introduces additional complexity, and may become challenging over heterogeneous land surfaces which violate the assumption of the model. By contrast, NIRvH1 and NIRvH2 completely avoid such additional complexity by taking the advantage of the features of soil and vegetation spectrum, and they show better performance than DVI and NIRv over sparse canopies (Fig. 5). NIRvH1 and NIRvH2 also performed better than DVI and NIRv over dense canopies when the soil has minimal impacts (Fig. S4), possibly due to that DVI (=NIR-Red) and NIRv (=NIR·NDVI) were not equal to NIR with the additional term of Red (\neq 0) and NDVI (\neq 1) in DVI and NIRv, respectively.

Table 2 summarizes the major characteristics of the new (NIRvH1 and NIRVH2) and traditional vegetation indices (DVI and NIRv) discussed in this study. NIRvH1 has the advantage that no assumption on the spectral shape of background soil is required and thus is more flexible. However, NIRvH1 needs the spectral information of the nearby pure soil pixels or at least a soil spectral database to drive the SVD approach. In addition, hyperspectral data are required which also limits the applicability of NIRvH1 in practice. NIRvH2 is simple to use and does not need any other additional information such as the nearby pure soil pixels or a soil spectral database. NIRvH2 also does not necessarily require hyperspectral data, as long as the slope k in Eq. 6 can be derived from a few red and NIR bands by multispectral sensors based on the assumption of a linearly increasing soil reflectance at the red edge. This assumption may not always be true but is reasonable for most soil types (Wang et al. 2017;

Yang et al. 2019). In addition, both NIRvH1 and NIRvH2 need to use reflectance measurement at 678 nm, the central wavelength of the chlorophyll absorption peak (Fig. S2a) which is the least sensitive to chlorophyll content and has the local minimal reflectance in the red band (Fig. 1). However, the central wavelength of the red band of many existing multispectral sensors is away from 678 nm (e.g., it is 648 nm for MODIS) in red. Cautions must be taken that NIRvH1 and NIRvH2 calculated with a red band departing from 678 nm may not have the best performance on removing soil impacts because of red band's sensitivity to the chlorophyll content (Fig. 1).

**Table 2** Major characteristics of the four vegetation indices: NIRvH1, NIRvH2, DVI and NIRv on soil impacts as candidates for NIR<sub>BS</sub>.

Remote sensing indices	Reduce the soil effect?	Influenced by the increase of soil reflectance?	Require hyperspectr al data?	Other characteristics
NIRvH1	Yes	No	Yes	Requires soil pixel or a soil spectral database for SVD; Narrow-band at 678 nm required
NIRvH2	Yes	No	Not necessarily	No additional information needed; Simple; Narrow- band at 678 nm required
DVI	Partially	Yes	No	Maximum offset on the SIF-VI relationship
NIRv	Partially	Yes	No	Medium offset on the SIF- VI relationship

## 5.2 Uncertainties and requirements in NIRvH estimations

Multiple scattering between the vegetation and soil, i.e., BRF<sub>M</sub> in Eq. 1, also contributes to the canopy total reflectance in both leaf spectral-invariant regions at 675~681 nm and 778~800 nm. In this study, BRF<sub>M</sub> has been partially included in NIRvH (NIRvH1 and NIRvH2) by the curve fitting, enabling a better correlation between NIRvH and SIF or NIR<sub>BS</sub> than the widely-

used NIRv (Fig. 5). However, the importance of including the multiple scattering contributions in NIRvH depends on the specific application scenarios. For example, if the research goal is to use NIRvH for estimating GPP, to normalize SIF and retrieve  $\Phi_F$ , or to correct SIF for viewangle effects, BRF<sub>M</sub> should be included in NIRvH, since soil reflected photons can be reabsorbed by the canopy, and then excite fluorescence and trigger photosynthesis. When applying NIRvH for retrieving canopy structural parameters (e.g., LAI, leaf inclination angle and clumping index), BRF<sub>M</sub> should be excluded in the calculation of the NIRvH, because all the soil contributions, no matter from single or multiple scattering, belong to the noise signal for estimating canopy structural parameters.

Leaf reflectance at 678 nm is small (usually <0.1 and mostly <0.05, Fig. 1) but a non-zero positive value, because non-photosynthetic biochemical constituents within the leaves, such as the dry matter and brown pigments, always scatter a small proportion of incident radiation at 678 nm. NIRvH2 not only reduces the soil contribution from the canopy total reflectance, but also reduces the scattering by non-chlorophyll leaf constituents. In fact, all scattering contributions by non-chlorophyll components can be reduced from BRF<sub>T</sub>(NIR) by NIRvH. In principle, NIRvH is robust and flexible for various application scenarios. As long as the background spectra of non-chlorophyll components (e.g., soil, litter, impervious surfaces and water body) can be identified in nearby pixels (for NIRvH1), or the slope of the background spectra varies minimally in the range of 675~800 nm (for NIRvH2), NIRvH is applicable over either natural or urban ecosystems (Fig. S3). Note that NIRvH can only separate the NIR contribution by chlorophyll and non-chlorophyll components, while cannot distinguish the chlorophyll contribution from leaves, woody parts (e.g., branches, stems and bark) and understory (e.g., moss and lichen). This allows NIRvH and SIF have similar radiative transfer process and results in the strong correlation between NIRvH with SIF (Figs. 5 and 10), although differences and uncertainties could come from the contribution of brown pigments that cause a slight positive slope in the NIR region of 778~800 nm and violates the assumption of flat canopy NIR reflectance (Fig. S1).

We recommended using two leaf spectral-invariant regions in the red and NIR bands for calculating NIRvH. In fact, the blue band at 450~480 nm is another spectral-invariant region that is not sensitive to canopy structure variations and potentially useful for calculating NIRvH. However, this may be challenging to use the blue band to calculate NIRvH, particularly NIRvH2 for two reasons. First, it might not be reasonable to assume soil reflectance linearly increases over a large spectral region from blue to red edge, and thus does not meet the prerequisite of the NIRvH2 algorithm. Second, atmospheric correction at the blue band is more complicated due to aerosol scattering and remaining larger artefacts than the red and NIR bands (Vermote et al., 1997). However, the NIRvH algorithm can still be promising to reduce the soil impacts of other vegetation indices at the canopy scale, such as the Photochemical Reflectance Index (PRI) (Gamon et al., 1992) and Chlorophyll/Carotenoid Index (CCI) (Gamon et al., 2016), which have typically been applied with airborne or ground-based measurements with limited atmospheric scattering impacts on the blue band.

#### 5.3 Potential applications of the hyperspectral-based NIRvH

NIRvH could be useful to decouple the soil and vegetation contribution in mixed pixels for current and forthcoming hyperspectral missions. TROPOMI covers 675~775 nm (Guanter et al., 2015) but not the largest chlorophyll absorption wavelength at 778 nm. However, applying both of our NIRvH approaches within the TROPOMI's spectral coverage range could still perform well, as indicated by SCOPE experiments (Figs. 6 and 9). The forthcoming FLEX mission covers a wider spectral range of 500~780 nm (Drusch et al., 2016). The new hyperspectral instrument German Aerospace Center (DLR) Earth Sensing Imaging Spectrometer (DESIS) onboard the International Space Station (ISS) covers the wavelength of

400~1000 nm with the spectral resolution of 2.5 nm (Krutz et al., 2019), which will be promising for further validating our NIRvH approaches especially by including the spectralinvariant NIR region of 778~800 nm. However, similar to other VIs for reducing soil impacts such as NDGI (Yang et al. 2019), NIRvH needs inputs from at least two red or NIR wavelengths/bands. This suggests that NIRvH designed for hyperspectral sensors is not applicable for most existing multispectral sensors/satellites such as MODIS and Landsat-8. Nevertheless, the NIRvH2 framework may be applicable with Sentinel-2 measurements which have 2 bands in NIR regions to partially reduce the soil impacts, although both Sentinel-2 NIR bands are beyond 800 nm and could introduce additional uncertainties by extrapolating Eq. 6. The reduced sensitivity of the NIRvH for soil background compared to the NIRv and its strong linear correlation to SIF enables it to approximate "potential SIF". This may open many important applications, including the normalization of SIF and obtaining canopy-scale SIF yield ( $\Phi_F$ ). Zeng et al (2019) demonstrated that SIF and NIRv radiance (Baldocchi et al., 2020; Wu et al., 2020) were related through their joint dependence on canopy structure and sunsensor geometry. The residual between the two measurements (NIRv or NIRvH radiance and SIF) should be directly related to  $\Phi_F$  (Wang et al., 2020). Variations in  $\Phi_F$  relate to the differential partitioning of absorbed PAR between photochemical and non-photochemical processes at the photosystem level. While the difference in performance between NIRvH and NIRv might be minor, small uncertainties could propagate directly into larger errors in  $\Phi_F$ . When combined with high-frequency SIF and eddy covariance measurements, NIRvH can be used to infer plant responses to environmental changes and stresses such as drought and thus improve our mechanistic understanding of the underlying physiological and ecological processes. Furthermore, since SIF measurements are usually sparse and coarse, NIRvH, as the "potential SIF", is a good candidate as the reference for estimating SIF at a higher spatiotemporal resolution. This would also allow us to look back in time and possibly extend SIF

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time series to the past when suited hyperspectral data (e.g., Hyperion) or applicable multispectral data were available.

Note that in addition to NIRv and DVI, some other vegetation indices have been proposed to reduce the soil impacts, such as the soil-adjusted vegetation index (SAVI) (Huete et al., 1988), enhanced vegetation index (EVI) (Huete et al., 2002), plant phenology index (PPI) (Jin and Eklundh, 2014), normalized difference phenology index (NDPI) (Wang et al., 2017) and normalized difference greenness index (NDGI) (Yang et al., 2019). They are widely used in the time-series analysis of phenology, but it is difficult to link their absolute magnitudes with the photon escape ratio (*fesc*) of SIF, and thus they cannot be directly used to calculate the total emitted SIF by directional observed SIF. Thus we mainly focus on the improvement of NIRv and DVI with hyperspectral dataset as a proxy of "potential SIF".

## 6. Conclusions

Our study demonstrates a considerable dependence on the widely used NIRv and DVI for soil background contributions that translate into uncertainties of downstream applications such as estimates of GPP or the retrieval of  $\Phi_F$ . We showed that a rigorous exploitation of spectral invariant regions in the red and NIR enables the design of new VIs (NIRvH1 and NIRvH2) that show substantially reduced sensitivity for soil background contributions and even enables decoupling the combined contributions from vegetation and soil. We recommend using hyperspectral data for the NIRvH retrieval since such data enable a more robust usage of spectral invariant wavelength regions to estimate the spectral shape of soil background reflectance and eventually compensate this contribution. The newly designed NIRvH can be considered as a robust proxy of "potential SIF" and enables various applications to contribute to ecological research. We suggest evaluating the potential of NIRvH to directly approximate GPP, to normalize SIF retrievals with structural effects and illumination effects for retrieval of

 $\Phi_F$ . The NIRvH also holds potential to fill gaps in sparse spatio-temporal sampling of actual SIF and could even be exploited to extend available SIF time series to the past. These fields of applications are hypothetical, and we recommend further detailed experiments to exploit these possibly highly interesting application fields.

## Acknowledgement

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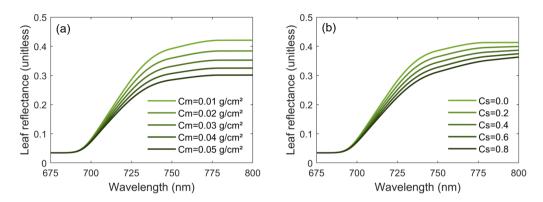
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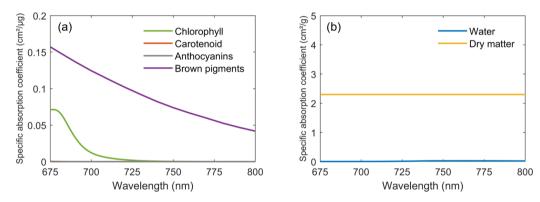
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# Supplementary Information



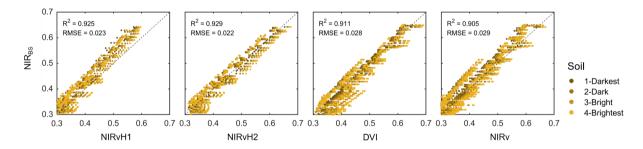
**Fig. S1** The leaf reflectance spectra simulated by the PROSPECT-D model with different dry matter contents (a) and senescent material fractions (b). Here, the chlorophyll content is  $60 \, \mu g \, cm^{-2}$ , the carotenoid content is  $20 \, \mu g \, cm^{-2}$ , the equivalent water thickness is  $0.009 \, cm$  and the leaf structure parameter N is 1.4.



**Fig. S2** Specific absorption coefficients of leaf spectrum for chlorophyll, carotenoid, anthocyanins and brown pigments (a), and for water and dry matter (b) from the recent version of PROSPECT-D model (Féret et al. 2017) with the spectral range of 675~800 nm at the red edge which we are interested in for the NIRvH estimation.



**Fig. S3** The four study regions with HyPlant imageries acquired on August 23<sup>rd</sup>, 2012 (Region A: 50°52'37.52"N/ 6°26'41.89"E, 2315 m×595 m), June 30<sup>th</sup>, 2015 (Region B: 50°52'33.42"N/ 6°26'10.83"E, 2115 m×1550 m), July 19<sup>th</sup>, 2016 (Region C: 50°52'43.95"N/ 6°26'11.79"E, 1265 m×1115 m), and August 23<sup>rd</sup>, 2012 (Region D: 50°54'39.10"N/ 6°26'41.89"E, 1950 m×455 m), respectively.



**Fig. S4** Scatter plots between NIR<sub>BS</sub> and different VIs by SCOPE simulations over dense canopies at different combinations of canopy structure, leaf property, sun-sensor geometry and soil spectrum in Table 1. To highlight the condition of dense vegetation, we examined the results where all the VIs were no less than 0.3.